Ok so briefly you will need that:

4% agarose in PBS to section brain

1) Stockage solution (for the brains just after dissection from the skull):

- 24h in ~~PFA~~ 4% / PBS at 4°C (not a big deal if you let them 48-72h !)

- Then stock in PBS + 0.1% ~~Sodium Azide~~ at 4°C (for long term in the fridge)

2) Cryobuffer (for the slices when you cut them and to stock them also in eppendorff or plates)

For 250 ml of cryobuffer:   - 250 ml ~~PBS~~

- 150 ml ~~ethylene glycol~~

- 100 ml ~~glycerol~~

- 0.5g Sodium Azide

3) You will need ~~3% H~~~~2~~~~O~~~~2~~ (from Sigma for example) : only needed for DAB staining (not flueorescence)

4) You will need ~~Triton x100~~ (for permeabilisation)

5) You will need also some serum (for the blocking solution). You can buy several like NGS (~~Normal Goat Serum~~). You can find some people said that Normal Serum that you use to block should be from the same specie than your secondary antibody. I never paid attention to it…

6) You will of course need antibodies:

- primary antibodies: the AT8 that we have in this lab is from Thermo Scientific (~~#MN1020~~)

- secondary antibodies:

if you do fluorescence one day you have to buy some Alexa fluorescent antibodies (I will show you in our lab)

if you do visible staining with DAB (like I did last time) you need to buy some secondary antibodies-biotinylated and then do the ABC kit and then the DAB kit (I will write that in the protocol).

* 2ndary for DAB: Because AT8 is a mouse if I remember you need a biotinylated anti-mouse (~~BA-2000 from the company VECTORLABS~~)
* The ~~ABC kit~~ we use the one from VECTASTAIN, PK-4000
* The ~~DAB kit~~ we use the one from KPL (DAB Reagent Set #54-10-00)

Of course you need also some Slides (~~Superfrost plus from Fisherbrand~~) and cover glass Microscope Cover Glass 22x50-1.5 from Fisherbrand) but we have some of that so I can give it to you.